

**Table III. Predominant pumps and carriers in the plasma membrane of guard cells and their functional characteristics**

Genetic codes relate to Arabidopsis; functional data relate to *V. faba* unless noted otherwise. For  $I_{sat}$ , current saturation is at  $V > > 0$  for ATPases or  $V < < 0$  for coupled transport;  $I_v$  are typical currents within the physiological voltage range; currents are converted where necessary assuming  $1 \mu A cm^{-2} = 1 pA pF^{-1}$ , guard cell surface area of  $3 \times 10^{-5} cm^2$ , and volume of 4 pL. Shorthand identifiers in parentheses cross-reference to Box 1. n.d., not determined. References are as follows: A (Blatt, 1987a; Blatt et al., 1990a; Lohse and Hedrich, 1992; Thiel et al., 1992; Becker et al., 1993; Gaxiola et al., 2007); B (Gräf and Weiler, 1990; Camelli et al., 1992; Askerlund, 1997; Palmgren and Harper, 1999; Bonza et al., 2000; Geisler et al., 2000; Sze et al., 2000; Bonza and De Michelis, 2011; Pittman, 2011); C (Rodríguez-Navarro et al., 1986; Blatt et al., 1987; Blatt and Slayman, 1987; Clint and Blatt, 1989; Maathuis and Sanders, 1994; Maathuis et al., 1997; Quintero and Blatt, 1997; Kim et al., 1998; Rubio et al., 2000, 2008; Chérel et al., 2002; Gierth et al., 2005; Nieves-Cordones et al., 2008; Remy et al., 2013); D (Beilby and Walker, 1981; Sanders and Hansen, 1981; Sanders et al., 1985, 1989; Meharg and Blatt, 1995; Guo et al., 2003; Hawkesford and Miller, 2004); E (Reddy and Das, 1986; Slone et al., 1991; Buckhout, 1994; Ritte et al., 1999; Stadler et al., 2003; Lee et al., 2008; Bates et al., 2012; Santelia and Lawson, 2016); F (Kang et al., 2010; Kuromori et al., 2010, 2011; Kanno et al., 2012; Zhang et al., 2014).

Transporter	Name	Locus	Function	No.	Stoichiometry	$I_{sat}$	$I_v$	$E_{rev}$	Ion Selectivity	References
				$\times 10^6 cell^{-1}$		$\mu A cm^{-2}$	$\mu A cm^{-2}$	mV		
H <sup>+</sup> -ATPase (H-ATPase)	AHA1 AHA2 AHA5	AT2G18960 AT4G30190 AT2G24520	H <sup>+</sup> extrusion, energization	1–3*	1 H <sup>+</sup> :1 ATP	22 ± 5 15 ± 5 <sup>a</sup>	2–15 5–9 <sup>a</sup>	–359 to –457 <sup>3,a,d</sup>	n.d.; assumed selective from pH dependence	A
Ca <sup>2+</sup> -ATPase (Ca-ATPase)	ACA1 ACA3 ACA8 ACA10 ACA11 ACA12	AT1G13210 AT1G27770 AT1G07810 AT5G57110 AT4G29900 AT3G57330 AT3G63380	Ca <sup>2+</sup> extrusion	0.1–0.3**	1 Ca <sup>2+</sup> :1 ATP	1–2	n.d.	–200 <sup>4</sup>	n.d.	B
H <sup>+</sup> /Ca <sup>2+</sup> antiport	CAX11	AT1G08960	Ca <sup>2+</sup> extrusion	0.1–0.3**	2 or 3 H <sup>+</sup> :1 Ca <sup>2+</sup>	n.d.	–0.6 <sup>1,b</sup>	>+100 <sup>5</sup>	Transport of K, Na, Mn, Zn, Li possible	B
H <sup>+</sup> -K <sup>+</sup> symport (H-K symport)	KUP/HAK/KT1 KUP/HAK/KT2 KUP3 = KT4 KUP/KT5 HAK5 KUP/HAK/KT6 KUP/HAK/KT7 KUP/HAK/KT8 KUP/HAK/KT9 KUP/HAK/KT10 KUP/HAK/KT11 ZIFL1.3	AT2G30070 AT2G40540 AT3G02050 AT4G33530 AT4G13420 AT1G70300 AT5G09400 AT5G14880 AT4G19960 AT1G31120 AT2G35060 AT5G13750	K <sup>+</sup> uptake	0.1–0.5**	1 H <sup>+</sup> :1 K <sup>+</sup>	–2 to –12 <sup>c</sup> –0.1 to –0.3 <sup>b</sup>	–0.08 to –0.3 –1 to –2 <sup>c</sup> –0.1 <sup>b</sup>	>0 <sup>6,c</sup> +22 <sup>7,b</sup>	K <sup>+</sup> ~ Rb <sup>+</sup> >> Cs <sup>+</sup>	C
H <sup>+</sup> -Cl <sup>–</sup> (NO <sub>3</sub> <sup>–</sup> ) symport (H-Cl symport)	NRT1.1 NRT2.1	AT1G12110 AT1G08090	Inorganic anion uptake	0.1–0.5**	2 H <sup>+</sup> :1 Cl <sup>–</sup> (NO <sub>3</sub> <sup>–</sup> )	–3 to –12 <sup>b</sup> –1 to 3 <sup>a</sup>	–1 to –2 <sup>b</sup> –0.4 to –1 <sup>b</sup>	Near 0 <sup>8,a,b</sup>	n.d.	D

(Table continues on following page.)

**Table III. (Continued from previous page.)**

Transporter	Name	Locus	Function	No.	Stoichiometry	$I_{sat}$	$I_v$	$E_{rev}$	Ion Selectivity	References
H <sup>+</sup> -Mal symport	ABCB14	AT1G28010	Malate uptake	n.d.	3 H <sup>+</sup> :1 Mal <sup>2–</sup> ***	n.d.	n.d.	>+20 <sup>9</sup>	Malate ~ fumarate >> succinate ~ citrate	E
H <sup>+</sup> -sugar symport	SUC1 SUC3 STP1 STP4	AT1G71880 AT2G02860 AT1G11260 AT3G19930	Sugar uptake	0.02–0.05**	1 H <sup>+</sup> :1 sugar***	n.d.	–0.01 to –0.03 <sup>2,6,f</sup>	>+20 <sup>10</sup>	Various hexose sugars	E
ABA transport	ABCG22 ABCG40 AIT1 ABCG25 DTX50	AT5G06530 AT1G15520 AT1G69850 AT1G71960 AT5G52050	ABA import ABA import ABA import ABA efflux ABA efflux	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	F

<sup>a</sup>*Chara corallina*. <sup>b</sup>Arabidopsis. <sup>c</sup>*Neurospora crassa*. <sup>d</sup>*Vicia faba*. <sup>e</sup>*Commelina communis*. <sup>f</sup>*Pisum sativum*. <sup>1</sup>Estimated for 3 H<sup>+</sup>:1 Ca<sup>2+</sup>. <sup>2</sup>1 mM sugar, pH<sub>o</sub> 5.5. <sup>3</sup>pH<sub>o</sub> 4.5 to 7.5. <sup>4</sup>1 mM [Ca<sup>2+</sup>]<sub>o</sub>. <sup>5</sup>3 H<sup>+</sup>:1 Ca<sup>2+</sup>, pH<sub>o</sub> 5.5, 1 mM [Ca<sup>2+</sup>]<sub>o</sub>. <sup>6</sup>200 μM K<sup>+</sup>, pH<sub>o</sub> 6.1. <sup>7</sup>275 μM K<sup>+</sup>, pH<sub>o</sub> 4.5. <sup>8</sup>pH<sub>o</sub> 7, 100 μM Cl<sup>–</sup>/NO<sub>3</sub><sup>–</sup>. <sup>9</sup>0.1 mM Mal, pH<sub>o</sub> 6.1. <sup>10</sup>0.1 mM Suc, pH<sub>o</sub> 6.1. \*Calculated assuming a transport rate of 60 H<sup>+</sup> s<sup>–1</sup> (Sze et al., 1999), a current of 15 to 20 μA cm<sup>–2</sup> (Blatt, 1987a; Clint and Blatt, 1989), and guard cell surface area of 3 × 10<sup>–5</sup> cm<sup>2</sup>. \*\*Calculated from  $I_{sat}$  or the typical transport current assuming a transport turnover rate of 50 s<sup>–1</sup> and guard cell surface area of 3 × 10<sup>–5</sup> cm<sup>2</sup>; estimates for the Ca<sup>2+</sup>-ATPase are based on assumption of a 30-fold lower density than the plasma membrane H<sup>+</sup>-ATPase (Sze et al., 2000) and H<sup>+</sup>-coupled transporters scaled accordingly by current densities. \*\*\*Stoichiometry determined as the minimum thermodynamic requirement to drive net accumulation.