Table S1. Metabolite pool sizes

Compound	LL	nmol·gFW ⁻¹				nmol⋅mg ⁻¹ chlorophyll	
		Ref. 5	Ref. 16	Ref. 17	HL-ACC	LL	HL-ACC
RUBP	71.0 ± 20.2	46.7 ± 8.2	118.0 ± 11.0	42.0 ± 8.2	10.2 ± 7.1	115.1 ± 32.7	25.9 ± 18.0
3-PGA	180.2 ± 33.9	200.0 ± 45.0	168.0 ± 15.0	n.d.	57.3 ± 45.6	292.5 ± 55.1	145.5 ± 115.8
DHAP	66.5 ± 18.1	2.7 ± 0.6	57.3 ± 4.2	13.2 ± 2.9	30.0 ± 12.4	108.0 ± 29.4	76.2 ± 31.5
FBP	11.64 ± 4.5	8.9 ± 2.3	31.2 ± 2.4	3.1 ± 0.2	3.6 ± 2.0	18.8 ± 7.3	9.2 ± 5.1
G6P	228.1 ± 56.5	173.0 ± 51.0	272.0 ± 15.0	159.2 ± 17.4	109.5 ± 35.3	370.2 ± 91.7	278.1 ± 89.5
F6P	175.0 ± 16.9	86.4 ± 14.6	128.0 ± 8.0	71.5 ± 12.5	126.4 ± 21.8	283.9 ± 27.5	321.0 ± 55.4
G1P	29.0 ± 12.7	11.7 ± 2.4	11.4 ± 1.2	17.9 ± 1.6	33.7 ± 7.9	47.0 ± 20.7	98.7 ± 20.0
S7P	58.9 ± 12.0	28.0 ± 5.4	87.5 ± 4.3	33.8 ± 11.6	48.4 ± 14.1	95.6 ± 19.5	122.9 ± 35.9
R5P	5.2 ± 0.8	1.2 ± 0.2	3.3 ± 0.8	6.2 ± 1.8	3.9 ± 1.0	8.5 ± 1.4	10.0 ± 2.5
UDPG	163.4 ± 38.7	35.7 ± 5.7	151.0 ± 4.0	86.0 ± 4.8	127.2 ± 37.2	265.1 ± 62.7	323.1 ± 94.4
ADPG	2.7 ± 0.7	0.6 ± 0.1	1.0 ± 0.1	1.0 ± 0.3	5.0 ± 1.9	4.5 ± 1.2	12.7 ± 4.9
Glycerate	101.1 ± 18.2	169.0 ± 65.0	290.0 ± 11.0	522.3 ± 101.0	209.4 ± 66.4	164.1 ± 29.6	531.8 ± 168.7
2-PGA	8.5 ± 3.4	20.0 ± 4.5	nd	nd	3.6 ± 2.2	13.8 ± 5.5	9.0 ± 5.7
Glycolate	33.1 ± 9.2	nd	nd	nd	44.8 ± 5.1	53.8 ± 14.9	113.8 ± 13.0
Aconitate	143.0 ± 61.9	14.5 ± 5.5	22.8 ± 1.6	nd	106.3 ± 40.5	232.0 ± 100.5	270.0 ± 102.9
2-OG	174.7 ± 27.1	63.1 ± 18.8	90.4 ± 2.6	132.6 ± 32.8	236.8 ± 87.5	283.4 ± 43.9	601.4 ± 222.2
Succinate	353.5 ± 105.4	84.0 ± 48.2	122.0 ± 7.0	nd	472.3 ± 201.0	573.6 ± 171.1	$1,199.4 \pm 510.4$
Fumarate	$12,988.8 \pm 1,620.8$	$1,154.0 \pm 47.0$	nd	nd	26,330.2 ± 3,621.6	21077.1 ± 2,630.2	$66,866.4 \pm 9,197.2$
Malate	9,213.0 ± 1,381.4	$1,820.0 \pm 547.0$	$3,222.0 \pm 185.0$	$1,1147.0 \pm 1,217.0$	9,301.1 ± 1,557.1	14,950.1 ± 2,241.7	23,620.5 ± 3,954.4

Metabolite pool sizes in LL and HL-ACC leaves were measured on the basis of leaf fresh weight and chlorophyll content (SD, n > 4), and compared with reported metabolite data (1-3). Although generally comparable several metabolite pools decreased in size with acclimation. The changes were further confirmed with independent replicates involving additional experiments that included spiked standards (Dataset S1). All samples were processed identically with care to avoid shading and using internal standards to correct for effects due to sample loss during the extraction, matrix contributions, and ion suppression. Of note, RUBP concentration dropped the most in the HL-ACC condition, although remained comparable to others' measured values (3–6). Hexose phosphate levels were far from equilibrium, consistent with a regulatory role for starch production relative to RuBP regeneration (7, 8). Other metabolites such as organic acids were comparable to literature values (3, 9, 10). Further raw data can be found in Dataset S1. nd, not detected.

- 1. Szecowka M, et al. (2013) Metabolic fluxes in an illuminated Arabidopsis rosette. Plant Cell 25(2):694–714.
- 2. Arrivault 5, et al. (2009) Use of reverse-phase liquid chromatography, linked to tandem mass spectrometry, to profile the Calvin cycle and other metabolic intermediates in Arabidopsis rosettes at different carbon dioxide concentrations. Plant J 59(5):826-839.
- Timm S, et al. (2012) Glycine decarboxylase controls photosynthesis and plant growth. FEBS Lett 586(20):3692–3697.
 Laber LJ, Latzko E, Gibbs M (1974) Photosynthetic path of carbon dioxide in spinach and corn leaves. J Biol Chem 249(11):3436–3441.
- 5. Bassham JA, Kirk M, Jensen RG (1968) Photosynthesis by isolated chloroplasts. I. Diffusion of labeled photosynthetic intermediates between isolated chloroplasts and suspending medium. Biochim Biophys Acta 153(1):211–218.
- 6. Heber U (1976) Energy coupling in chloroplasts. J Bioenerg Biomembr 8(3):157–172.
 7. Schleucher J, Vanderveer P, Markley JL, Sharkey TD (1999) Intramolecular deuterium distributions reveal disequilibrium of chloroplast phosphoglucose isomerase. Plant Cell Environ 22(5):525-533.
- 8. Dietz K-J (1985) A possible rate-limiting function of chloroplast hexosemonophosphate isomerase in starch synthesis of leaves. BBA/Gen Subjects 839(3):240-248.
- 9. Pracharoenwattana I, et al. (2010) Arabidopsis has a cytosolic fumarase required for the massive allocation of photosynthate into fumaric acid and for rapid plant growth on high nitrogen. Plant J 62(5):785-795.
- 10. Araújo WL, et al. (2012) Antisense inhibition of the 2-oxoglutarate dehydrogenase complex in tomato demonstrates its importance for plant respiration and during leaf senescence and fruit maturation. Plant Cell 24(6):2328–2351.
- 16. Grafahrend-Belau E, et al. (2013) Multiscale metabolic modeling: Dynamic flux balance analysis on a whole-plant scale. Plant Physiol 163(2):637-647.
- 17. Borisjuk L, et al. (2013) Seed architecture shapes embryo metabolism in oilseed rape. Plant Cell 25(5):1625-1640.