

Table 1
Effects of crowders on protein stability.

Protein	Crowder	Crowder concentration	Experimental conditions	Observations	References
FKBP (FK-506 binding protein)	Dextran 6, 10, 20, 40, 70, 100 and 150 kDa Ficoll 70 kDa	0–200 g l ⁻¹	21.5 °C and pH 6.5	Maximum stabilization at 40 kDa (100 g l ⁻¹). The difference in free energy change, $\Delta\Delta G (\Delta G_{D40}^a - \Delta G_{buffer}^b) = 0.59 \text{ kcal mol}^{-1}$, where $\Delta G_{D40} = 5.35 \text{ kcal mol}^{-1}$ and $\Delta G_{buffer} = 4.76 \text{ kcal mol}^{-1}$.	[30]
ϵ and θ subunits of the <i>E. coli</i> DNA polymerase III holoenzyme (Pol III)	Dextran 6, 40, 70, 100 and 150 kDa Ficoll 70 kDa	100 g l ⁻¹	15 °C and pH 7.6	Increase in binding constant of ϵ 186 and θ subunits in the presence of dextran but it diminished with increasing size of dextran suggesting decrease in stabilizing effect. The binding constants in buffer and dextran 6 were 1.4 and 7.5 μM^{-1} , respectively.	[50]
Horse heart cytochrome c	Dextran 40 and 70 kDa Ficoll 70 kDa	0–400 g l ⁻¹	20 °C and pH 7.0	Dextran 40 was more stabilizing than dextran 70 in the presence of GdmCl. $\Delta\Delta G_{ND}$ (the protein stability change due to crowding) from the simulations increased as the crowder size decreased. The changes in T_m were 5–10 °C in the case of ficoll 70 and dextran 70 and 10–20 °C for dextran 40, respectively.	[51]
HSA (human serum albumin)	Dextran 6, 40 and 70 kDa	0–175 g l ⁻¹	pH 7.0	Dextran 6 had the largest value of difference in urea-induced unfolding free energy change ($\Delta\Delta G^\circ = 12 \text{ kJ mol}^{-1}$) followed by dextran 70 ($\Delta\Delta G^\circ = 9.6 \text{ kJ mol}^{-1}$) and dextran 40 ($\Delta\Delta G^\circ = 5.2 \text{ kJ mol}^{-1}$) at 150 g l ⁻¹ .	[60]
<i>P. aeruginosa</i> apoazurin	Dextran 20, 40 and 70 kDa	0–300 g l ⁻¹	20 °C and pH 7.0	The size of the dextran did not matter for its contribution in the stability of apoazurin. The unfolding free energy changes (ΔG_{D0}°) were 36 and 40.2 kJ mol ⁻¹ in buffer and in all the three crowders (at 300 g l ⁻¹), respectively.	[52]
<i>A. vinelandii</i> apoflavodoxin	Dextran 6, 20 and 70 kDa	40% (w/v)	pH 6.0	Comparable increase in the T_m of apoflavodoxin unfolding in all the sizes of dextran. The changes in T_m ($\Delta T_m = T_{m,crowder}^c - T_{m,buffer}^d$) were 4.2, 3.8 and 4.2 °C for dextran 6, 20 and 70, respectively.	[53]
Equine skeletal muscle Myoglobin	Dextran 40 and 70 kDa Ficoll 70 kDa	0–200 g l ⁻¹	pH 7.0	Both the sizes of dextran showed slight variation than ficoll 70 with urea concentration in terms of T_m . The values of T_m decreased from 85.2 to 83.4 °C in 0 M urea and from 58.2 to 56.2 °C in 5 M urea, respectively in the case of ficoll 70. However, dextran 40 showed higher average fluorescence lifetime indicating greater Trp-heme separation than the other crowder.	[55]
c-MYC i-motif	PEG 0.2, 0.4, 1, 2, 4 and 8 kDa	20% and 40% (w/w)	pH 4.0–7.0	The T_m was dependent on pH and the molecular mass of PEG. At pH values above 4.5, the stabilization of the i-motif was increased with the increasing size of PEG. The maximum changes in T_m ($\Delta T_m = T_{m,PEG} - T_{m,buffer}^d$) were 15.8 °C for PEG 2 (20% w/w) and 27.4 °C for PEG 8 (40% w/w), respectively at pH 6.0.	[54]
Recombinant bovine odorant binding protein (bOBP)	PEG 0.6, 4 and 12 kDa	0, 80, 150 and 300 g l ⁻¹	pH 7.8	PEG 0.6 possessed more pronounced stabilizing effects than PEG 4 and 12 at moderate concentration, whereas at higher concentration, both PEG 0.6 and 4 stabilized more than PEG 12. The unfolding free energies (G_{N-D}°) were -7.7, -9.9, -8.3 and -9.1 kJ mol ⁻¹ for buffer, PEG 0.6 (300 g l ⁻¹), 4 and 12 (150 g l ⁻¹), respectively.	[71]

^a ΔG_{D40} represents ΔG in the presence of dextran 40.

^b ΔG_{buffer} represents ΔG in the absence of crowding agents.

^c $T_{m,crowder}$ and $T_{m,PEG}$ represent T_m in the presence of any crowding agent and PEG, respectively.

^d $T_{m,buffer}$ represents T_m in the absence of crowding agents.

- [30] J. Batra, K. Xu, H.X. Zhou, Nonadditive effects of mixed crowding on protein stability, *Proteins* 77 (2009) 133–138.
- [50] J. Batra, K. Xu, S. Qin, H.-X. Zhou, Effect of macromolecular crowding on protein binding stability: modest stabilization and significant biological consequences, *Biophys. J.* 97 (2009) 906–911.
- [51] A. Christiansen, Q. Wang, A. Samiotakis, M.S. Cheung, P. Wittung-Stafshede, Factors defining effects of macromolecular crowding on protein stability: an in vitro/in silico case study using cytochrome c, *Biochemistry* 49 (2010) 6519–6530.
- [52] A. Christiansen, P. Wittung-Stafshede, Quantification of excluded volume effects on the folding landscape of *Pseudomonas aeruginosa* apoazurin in vitro, *Biophys. J.* 105 (2013) 1689–1699.
- [53] R. Engel, A.H. Westphal, D.H. Huberts, S.M. Nabuurs, S. Lindhoud, A.J. Visser, C.P. van Mierlo, Macromolecular crowding compacts unfolded apoflavodoxin and causes severe aggregation of the off-pathway intermediate during apoflavodoxin folding, *J. Biol. Chem.* 283 (2008) 27383–27394.
- [54] J. Cui, P. Waltman, V. Le, E. Lewis, The Effect of molecular crowding on the stability of human c-MYC promoter sequence I-motif at neutral pH, *Molecules* 18 (2013) 12751.
- [55] A. Malik, J. Kundu, S.K. Mukherjee, P.K. Chowdhury, Myoglobin unfolding in crowding and confinement, *J. Phys. Chem. B* 116 (2012) 12895–12904.
- [60] S. Biswas, P.K. Chowdhury, Unusual domain movement in a multidomain protein in the presence of macromolecular crowders, *Phys. Chem. Chem. Phys.* 17 (2015) 19820–19833.

- [71] O.V. Stepananko, D.O. Roginskii, O.V. Stepanenko, I.M. Kuznetsova, V.N. Uversky, K.K. Turoverov, Structure and stability of recombinant bovine odorant-binding protein: III. Peculiarities of the wild type bOBP unfolding in crowded milieu, *PeerJ* 4 (2016) e1642.