Ecological and Physiological Studies on Purple Sulphur Bacteria (Chromatiaceae) at Aswan High Dam Lake

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Abstract: 322 isolates of purple sulphur bacteria were isolated from different localities of study in Aswan High Dam Lake. These comprise five genera of purple sulphur bacteria (*Chromatiaceae*). *Allochromatium* was the most common genus 161 (50%) followed by *Thiocystis 50* (15.53%), *Thiocapsa 40* (12.42%), *Thiodictyon* 38 (11.8%) and *Lamprocystis 33* (10.25%) from the total number of isolates during the whole period of study. These genera represented by six speces *Allochromatium vinosum* and *A. warmingii*; *Thiocystis violacea*; *Thiocapsa roseopersicina*; *Thiodictyon elegans and Lamprocystis roseopersicina*. Physico-chemical parameters of water samples were determined.

Key words:Purple sulphur bacteria · Allochromatium vinosum · A. warmingii · Thiocystis violacea · Thiocapsa roseopersicina · Thiodictyon elegans · Lamprocystis roseopersicina

INTRODUCTION

The water areas in Egypt include the River Nile and Aswan High Dam Lake (AHDL) previously referred to as Lake Nasser. The purple sulfur bacteria included in the gamma subdivision of proteobacteria capable of photosynthesis [1]. They are Gram negative found in illuminated anoxic zones of lakes and other aquatic habitats where hydrogen sulfide accumulates and not use water as their reducing agent and so do not produce oxygen. Instead they use hydrogen sulfide, which is oxidized to produce granules of elemental sulphur S inside the cells. The combination of the yellow and red carotenoids with the bacteriochlorophylls a or b form colors of red, brown, purple and sometimes orange.

In the traditional classification, morphological properties such as cell form and size, motility, presence of gas vesicles and formation of cell aggregates were properties of major relevance. Because this classification was not in congruence with the phylogenetic relationships within the Chromatiaceae species, a reclassification was proposed on the basis of 16S rDNA sequence similarities and with support of selected phenotypic properties [2, 3].

A number of more recent reviews consider ecological aspects and the distribution of purple sulfur bacteria in nature [4-15]. The most important properties that

determine the development and distribution of phototrophic bacteria in nature are the availability of light, the light intensity and the concentrations of sulfide and oxygen, which form countercurrent gradients in most of the natural habitats of purple sulfur bacteria .

The objective of the present work is to isolate for the first time the photosynthetic purple sulphur bacteria (*Chromatiaceae*) from fresh water of two sites Khor-El Ramla (site I) and Khor Manam (site II) in Aswan High Dam Lake based on phenotypic properties.

MATERIALS AND METHODS

Sampling: Fresh water samples were collected from Aswan High Dam Lake at two successive sites (khor-El-Ramla (I) to the west and khor-Manam (II) to the east). Physico-chemical characteristics: temperature, pH values, dissolved oxygen, sulphate, Chlorophyll-a and bacteriochlorophyll-a of water samples were determined according to Pfennig [5], Jackson [16] and APHA [17].

Enrichment Medium: Samples from different depths and localities were inoculated into white screw-capped bottles containing an enrichment medium [18, 19]. Liquid medium was sterilized in 30 ml white screw-capped bottles. They were filled with a proper volume and autoclaved for about 15 minutes at 120°C. The enrichment media were

inoculated with water and sediment samples and then exposed to tungsten light source (light intensity, 100-300 lux) at 25-35°C. After 7 to 10 days, the colours which indicate the presence of purple sulphur bacteria were observed. Pure cultures were obtained by repeated subculturing in agar shake tubes [20]. Isolates were identified to species level microscopy, pigmentation and physiological characters according to the scheme of J. Imhoff [21].

RESULTS AND DISCUSSION

Physico-chemical Parameters: During summer and autumn seasons water temperature decreased with depth from the surface to the bottom 32.1-15.5°C. This may be due to thermal stratification [22-26]. The pH values ranged from a minimum of 7.2 at the bottom of site II to maximum of 8.80 at the surface of site I in spring 2002. This may be due to the photosynthetic activity especially macrophytes [25-28].

Dissolved oxygen (mg/l) ranged from 8.08 at the surface of site II in winter to 0.25 at the bottom of site I in spring 2002. This may be attributed to photosynthetic production of oxygen by phytoplankton blooms [29-32]. The relative decrease in sulfate contents recorded at the bottom of the lake may be due to the reduction of sulfate (SO₄²) to sulfides (S²) by the action of sulfate reducing bacteria. Similar, the decomposition of organic matter usually produces anaerobic conditions leading to the formation of hydrogen sulfide [27,33].

Chlorophyll-a and Bacteriochlorophyll-a: chlorophyll-a ($\mu g \, l^{-1}$) content ranged from 4.3-12 at the bottom of site II in autumn 2001 to a maximum value at the surface of site I in summer 2001, while bacteriochlorophyll-a ($\mu g/l$) increased from bottom to surface 2.5-9.9. In General, chlorophyll-a decreased with increasing depth, while bacteriochlorophyll-a ($\mu g/l$) increased. The changes in chlorophyll-a were found to be correlated to the phytoplankton density [28, 29, 34, 35]; while bacteriochlorophyll-a due to the presence of photosynthetic bacteria [5, 28, 29, 36, 37].

Characterization and Identification of the Purified Isolates: 322 isolates of purple sulfur bacteria were obtained from different localities of study. These comprise five genera of purple sulfur bacteria (*Chromatiaceae*) with six different species. The characteristics of *Chromatiaceae* were shown in Table 1 and Fig. 1.

Allochromatium vinosum: Cells are rod shaped 2 and 2.5-6 μm, occasionally longer. Globules of S evenly distributed within the cell color of growing cultures, first yellowish to orange-brown, later brownish red. Photosynthetic pigments are bacteriochlorophyll a and carotenoids of the normal spirilloxanthin group.

Grow phototrophically under anoxic conditions in the light. Photosynthetic electron donors: sulfide, sulfur, thiosulfate, sulfite, molecular hydrogen, formate, acetate, propionate, pyruvate, fumarate, malate and succinate. Some strains utilize butyrate. Not utilized: sugars, sugar alcohols, alcohols, benzoate, citrate and amino acids. Nitrogen sources: ammonium salts and N2. Mesophilic freshwater bacterium with optimum growth at 25-35°C and pH range 6.5-7.6. Habitat: ponds and lakes with stagnant freshwater, sewage lagoons, brackish waters, estuaries, salt marshes and marine habitats containing hydrogen sulfide and exposed to light. Among the most widely occurring species of the family *Chromatiaceae*. The mol% G_C of the DNA is: 61.3-66.3 (Bd).

Allochromatium warmingii: Cells ovoid to rod shaped, 3.5-4.0 and 5-11 µm, sometimes longer, motile by a flagellar tuft which is usually 1.5-2 times the cell length and visible by bright field or phase contrast microscopy. In the presence of sulfide and light, globules of S are predominantly located at the two poles of the cell. Color of individual cells is grayish to slightly pink, color of cell suspensions pinkish to purple-violet. Photosynthetic pigments are bacteriochlorophyll a and carotenoids of the rhodopinal group. Strictly anaerobic and obligately phototrophic. Sulfide reduced media required. Sulfide and S used as photosynthetic electron donors. In the presence of sulfide and bicarbonate, acetate and pyruvate are photoassimilated. Not utilized: thiosulfate, sugars, alcohols, higher fatty acids, amino acids, benzoate, formate and most intermediates of the tricarboxylic acid cycle. Assimilatory sulfate reduction is absent. Nitrogen sources: ammonium salts, urea and N2. Vitamin B12 required. Freshwater bacterium with optimum growth at 25-30°C and pH range 6.5-7.3. Habitat: ditches, ponds and lakes with stagnant freshwater containing hydrogen sulfide and exposed to light. The mol% G C of the DNA is: 55.1-60.2

Lamprocystis Schroeter 1886, Brilliant Bag: Lamprocystis roseopersicina is distantly related to Thiocapsa and Thiodictyon species [38]. Sequence analyses of 16S rDNA place this species into the group of

Table 1: Characterization of Chromatiaceae isolated from two sites (Khor E-Ramla and Khor Manam) from Aswan High Dam Lake

	Genera and Species					
<u> </u>	Allochromatium	Allochromatium	Thiocystis	Thioapsa	Thiodictyon	Lamprocystis
Feature	vinosum	warmingii	violacea	roseopersicina	elegans	roseopersicina
Bacteriochlorophyll	7007	222	222	192022	10201	192022
absorption maxima	375	375	375	375	375	375
	590	590	559	590	590	590
	805	805	805	805	805	805
	830	830	830	830	830	830
	890	890	890	890	890	890
Bacteriochlorophyll	a	a	a	а	а	а
present						
Carotenoid	425	S. Const.	- 	35. Second	ia Carrier	250 mg
absorption maxima	460	465	470	485	470	464
	490	490	490	515	S S	490
	525	525	523	545	525	72
Carotenoid present	spirillo-	rhodopinal	okenone	spirillo-	rhodopinal	lycopinal
	xanthin	and	and	xanthin	and	and lycopinol
		rhodopinol	rho do pin al		rhodopin	purple-
Culture pigmentation	orange-	purple-	purple-	pink	purple-	violet
	brown	violet	violet		violet	spherical
Morphology	ovoid	ovoid to rod	spherical	spherical	rod shaped	to ovoid
Cell size (µm)		88/87/0				2.0 - 3.5
width	2	3.5-4	2.5-3.0	1.5	1.5 - 2	0547 S242
length	2.5-6	5-11		525	3.8	6.5 - 7.3
pH range	6.5-7.5	6.5 - 7.3	6.5-7.6		6.7-7.3	25-30°C
Temperature range	30-35°C	30-35°C	25-35°C		25-30°C	U55456 B
Photo assimilation:		4.01-5.000x				+
Acetate	19	Ť	+	+	+	+
Pyruvate	+	+	+	+	+	121
Furnarate	+	8	81	+	12	723
Malate	+	-	-	+	:-	n=1
Succinate	+	=	-	+	12	1901
Formate	(+)	*** ***	2	325	92 52	121
propionate	(+)	8	8	(2)	12	623
Glycerol	(S			+		N a 8
Fructose	15	© =	0	+	25 12	+
Sulphide	+	+	+	(+)	+	325
Thiosulphate	Ŧ	· · · · · · · · · · · · · · · · · · ·	¥		12	+
Bicarbonate	19	+	į.	+	+	18
Nitrogen souece:	26	C.	4.	200	58	+
ammonium salts	+	+	+	+	+	2000 2026
urea	12	¥	¥	(2)	12	

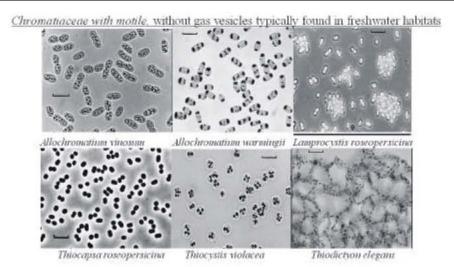


Fig. 1: Chromatiaceae isolated from two sites (Khor E-Ramla and Khor Manam) from Aswan High Dam Lake

spherical to ovoid, non motile freshwater *Chromatiaceae* species, many of which contain gas vesicles and in close proximity to the species originally described as *Amoebobacter purpureus* [39]. In accordance with the rules of the International Code of Nomenclature of Bacteria, the transfer of this species to the genus *Lamprocystis* as the new combination *Lamprocystis purpurea* comb.nov. was proposed [40].

Lamprocystis roseopersicina: Rosy peach-colored. Cells spherical to ovoid, 2.0-3.5 µm in diameter motile by flagella, contain gas vesicles. Color of cell suspensions pinkish violet to purple-violet. Photosynthetic pigments are bacteriochlorophyll a and carotenoids of the rhodopinal group, with lycopenal as main component [18]. Obligately phototrophic and strictly anaerobic. Photolithoautotrophic growth with sulfide and S as electron donors. In the presence of sulfide and bicarbonate, acetate and pyruvate are photoassimilated. Nitrogen sources: ammonium salts. Mesophilic freshwater bacterium with optimum growth at 20-30°C and pH 7.0-7.3. Habitat: mud and stagnant water of ponds and lakes containing hydrogen sulfide. The mol% G C of the DNA is: 63.8 (Bd). [40], 1700VP (Pfennigia purpurea [38, 41], 1308; Amoebobacter purpureus [39], 399.

Thiocapsa roseopersicina: Rosy peach-colored. Cells spherical, 1.2-3.0 µm, usually 1.5 µm in diameter and surrounded by a strong slime capsule. Aggregates of two, four, or more cells common; irregular clumps of cells usually surrounded by slime. Gas vesicles absent. Individual cells colorless, color of cell suspensions pink rose-red. Photosynthetic pigments bacteriochlorophyll a and carotenoids of the normal spirilloxanthin group [42, 43]. Photolithoautotrophic growth under anoxic conditions in the light with hydrogen, sulfide, thiosulfate and sulfur as electron donors. Acetate, pyruvate, fumarate, malate, succinate, glycerol and fructose are photoassimilated. Not utilized: lactate, propionate, butyrate, tartrate, citrate, benzoate and alcohols. Chemoautotrophic or chemoheterotrophic growth under microoxic to oxic conditions in the dark is possible. Most strains capable of assimilatory sulfate reduction. Nitrogen sources: ammonium Nitrogenase activity was present. Growth factors not required. Mesophilic bacterium with optimum growth at 20-35°C and pH range 6.5-7.5. Habitat: stagnant water and mud of ponds, pools or wastewater lagoons containing degradable organic substances and hydrogen sulfide.

Also common in estuaries and salt marshes. The mol% G C of the DNA is: 63.3-66.3

Thiocystis violacea: Violet-colored. Cells spherical and about 2.5-3 µm in diameter. Depending on the culture conditions, larger individual cells may occur. Irregular cell aggregates surrounded by slime are formed under unfavorable conditions. Sulfur globules appear randomly distributed within the cell. Color of individual cells grayish, color of cell suspensions purple-violet. Photosynthetic pigments are bacteriochlorophyll a and carotenoids of the rhodopinal group [42, 43]. Photoautotrophic growth occurs under anoxic conditions in the light with sulfide, sulfur, thiosulfate, sulfite (used by some strains) and molecular hydrogen as electron donors. In the presence of sulfide and bicarbonate, acetate, pyruvate and fumarate are photoassimilated. In addition, some strains use propionate, succinate, oxoglutarate, glucose, fructose, propanol or glycerol. Chemoautotrophic or mixotrophic growth possible under microoxic to oxic conditions in the dark. Some strains capable of assimilatory sulfate reduction. Nitrogen sources: ammonium salts, urea and N2. Mesophilic bacterium with optimum growth at 25-35°C and pH range 6.5-7.6. Marine isolates may tolerate or require low concentrations of NaCl.

Habitat: ponds and lakes with stagnant freshwater, brackish water, or seawater containing hydrogen sulfide; sewage lagoons, estuaries, salt marshes and sulfur springs. The mol% G C of the DNA is: 62.8-67.9

Thiodictyon elegans: Cells are rod shaped 1.5-2.0 and 3-8 µm grow in the form of irregular netlike aggregates, dependent on the sulfide concentration and light intensity. In cultures of higher population densities, cell chains break apart and the cultures contain predominantly single cells. Color of single cells grayish, color of cell suspensions light violet to purple-violet. Photosynthetic pigments are bacteriochlorophyll a and carotenoids of the rhodopinal group, with rhodopinal and rhodopin as major components [18]. Obligately phototrophic and strictly anaerobic. Photolithoautotrophic growth under anoxic conditions in the light with sulfide and S as electron donors. During oxidation of sulfide, S is transiently stored in the peripheral part of the cell that is free of gas vesicles. In the presence of sulfide and bicarbonate, acetate and pyruvate are photoassimilated. Nitrogen sources: ammonium salts. Mesophilic freshwater bacterium with optimum growth at 20-25°C, pH range 6.7-7.3. The mol% G C of the DNA is: 65.3-66.3.

REFERENCES

- Woese, C.R., W.G. Weisburg, C.M. Hahn, B.J. Paster, L.B. Zablen, B.J. Lewis, T.J. Macke, W. Ludwig and E. Stackebrandt, 1985. The phylogeny of purple bacteria: The gamma subdivision. Syst. Appl. Microbiol., 6: 25-33.
- Guyoneaud, R., J. Su ling, R. Petri, R. Matheron, P. Caumette, N. Pfennigand J.F. Imhoff, 1998. Taxonomic rearrangments of the genera *Thiocapsa* and *Amoebobacter* on the basis of 16S rRNA sequence analysis and description of *Thiolamprovum* gen. nov. Int. J. Syst. Bacteriol., 48: 957-964.
- Imhoff, J.F., J. Süling and R. Petri, 1998. Phylogenetic relationships among the Chromatiaceae, their taxonomic reclassification and description of the new genera Allochromatium, Halochromatium, Isochromatium, Marichromatium, Thiococcus, Thiohalocapsa and Thermochromatium. Int. J. Syst. Bacteriol., 48:1129-1143.
- Kondratieva, E.N., 1965. Photosynthetic Bacteria. Program for Scientific Translations. Jerusalem, Israel. Pfennig, N. 1967. Photosynthetic bacteria. Ann. Rev. Microbiol., 21: 285-324.
- 5. Pfennig, N., 1967. Photosynthetic bacteria. Ann. Rev. Microbiol., 21: 285-324.
- Pfennig, N., 1989a. Ecology of phototrophic purple and green sulfur bacteria. In: H.G. Schlegel and B. Bowien (Eds.) Autotrophic Bacteria. Springer-Verlag. New York, NY.
- Gorlenko, V.M., G.A. Dubinina and S.I. Kusnetsov. 1983. The ecology of aquatic microorganisms [monograph]. In: W. Ohle (Ed.) Binnengewasser. Schweizerbartsche Verlagsbuchhandlung. Stuttgart, Germany, pp. 254.
- Van Gemerden, H. and H.H. Beeftink, 1983. Ecology of phototrophic bacteria. In: J.G. Ormerod (Ed.). The phototrophic Bacteria: Anaerobic Life in the Light. Blackwell Science Publishing. Oxford, UK., pp: 146-185.
- Lindholm, T., 1987. Ecology of photosynthetic prokaryotes with special reference to meromictic lakes and coastal lagoons. ABO Academy Press. Abo, Finland.
- Imhoff, J.F., 1988a. Anoxygenic phototrophic bacteria. In: B. Austin (Ed.) Methods in Aquatic Bacteriology. John Wiley. Chichester, UK. 207-240.

- Imhoff, J.F., 1988b. Halophilic phototrophic bacteria.
 In: F. Rodriguez-Valera (Ed.) Halophilic Bacteria. CRC Press. Boca Raton, FL., pp: 85-108.
- Imhoff, J.F., 1992. Taxonomy, phylogeny and general ecology of anoxygenic phototrophic bacteria. In: N. G. Carr and N.H. Mann (Eds.) Biotechnology Handbook Photosynthetic Prokaryotes. Plenum Press. London, UK, pp: 53-92.
- Imhoff, J.F., 2001a. True marine and halophilic anoxygenic phototrophic bacteria. Arch. Microbiol., 176: 243-254.
- Madigan, M.T., 1988. Microbiology, physiology and ecology of phototrophic bacteria. In: A.J.B. Zehnder (Ed.) Biology of Anaerobic Microorganisms. John Wiley. Chichester, UK, pp: 39-111.
- Van Gemerden, H. and J. Mas, 1995. Ecology of phototrophic sulfur bacteria. In: Anoxygenic Photosynthetic Bacteria R.E. Blankenship, M.T. Madigan and C.E. Bauer (Eds.). Kluwer Academic Publishers. Dordrecht, The Netherlands, pp. 49-85.
- 16. Jackson, M.L., 1958. Soil Chemical Analysis. Constable, London.
- APHA, 1998. Standard methods for examination of water and waste water (20th Edn.), American Puplic Health Association, New York.
- Van Niel, C.B., 1971. Techniques for the enrichment, isolation and maintenance of photosynthetic bacteria.
 In: S.P. Collowick and N.V. Kaplan (Eds.) Methods in Enzymology. Academic Press. New York, NY. 23, part A:3-28.
- Pfennig, N., Markham, M.C. and Liaaen-Jensen, S. 1968. Carotenoids of *Thiorhodaceae*. 8. Isolation and characterization of a *Thiothece*, *Lamprocystis* and *Thiodictyon* strain and their carotenoids pigments. Arch. Mikrobiol., 62: 178-191.
- Van Niel, C.B., 1931. On the morphology and physiology of the purple and green sulfur bacteria. Arch. Microbiol., 3: 1-112.
- Bergey's Manual of Systematic Bacteriology Second Edition Volume Two The *Proteobacteria* Part B The *Gammaproteobacteria* (2005) Don J. Brenner Noel R. Krieg James T. Staley, pp. 3-37.
- Zaghloul, F.A., 1985. Seasonal variations of plankton in Lake Nasser Ph. D. Thesis, Faculty of Science, Suez Canal University, Suez, Egypt.
- 23. Iskaros, I.A., 1993. Ecological studies on the distribution of zooplankton and benthic fauna in Khor Kalabsha in Lake Nasser. Ph.D. Thesis, Faculty of Science, El-Mansoura University, El-Mansoura, Egypt.

- Mageed, A.A., 1995. Studies on zooplankton from Lake Nasser, Ph.D. thesis, Faculty of Science, Al-Azhar University, Cairo.
- El-Shabrawy, G.M. and H.J. Dumont, 2003. Spatial and seasonal variation of the zooplankton in the coastal zone and main khors of Lake Nasser (Egypt). Hydrobiologia, 491: 119-132.
- Ali, M.M., A.A. Mageed and M. Heikal, 2007. Importance of aquatic macrophyte for invertebrate diversity in large subtropical reservoir. Limnologica, 37: 155-169.
- Hann, B.J., 1995. Invertebrates associations with submersed aquatic plants in a prairie wetland. USF (Delta Marsh) annual report, Vol. 30.
- Mageed, A.A. and M.T. Heikal, 2006. Factors affecting seasonal patterns in epilimnion zooplankton community in one of the largest man-made Lakes in Africa (Lake Nasser, Egypt). Limnologica, 36: 91-97.
- El-Otify, A.M., 1985. Studies on phytoplankton of Aswan High Dam Lake. M. Sc. Thesis, Botany Dept. Fac. Sci., Aswan. Assiut University, pp. 140.
- Shoreit, A.A.M., A.M. Ahmed and M.S.A. Shabeb, 1989. Field and laboratory studies on some photosynthetic bacteria in Aswan High Dam Lake 1. Some physical and chemical characteristics. Int. Revue Ges. Hydrobiol., 74 (4): 419-432.
- Cheruvelil, K.S., P.A. Sorannoand and J. Madsen, 2001. Epiphytic macroinvertebrates along gradient of Eurasian watermilfoli cover. J. Aquatic Plant Manage., 39: 67-72.
- Cheruvelil, K.S., P.A. Soranno, J.D. Madsen and M.J. Roberson, 2002. Plant architecture and epiphytic macroinvertebrate communities: The role of an exotic dissected macrophyte. J. North American Benthol. Soc., 21(2): 261-277.
- Sakuma, M., T. Hanazatoand R. Nakazato, 2002. Methods for quantitative samlping of epiphytic microinvertebrates in Lake vegetation. Limnology, 3: 115-119.
- Zidan, M.A., 1983. Laboratory and field studies on Nile phytoplankton, Ph.D. Thesis, Assiut University, Egypt, pp. 185.

- Ahmed, Z.A., 1983. Studies on phytoplankton of the Nile system in Upper Egypt. M.Sc. Thesis Assiut University, Egypt, pp: 144.
- Van Niel, C.B., 1944. The culture, general physiology, morphology and classification of the non-sulphur purple and brown bacteria. Bacteriol. Rev., 8: 1-118.
- 37. Herbert, R.A., 1976. Isolation and Identification of photosynthetic bacteria (*Rhodosirillaceae*) from Antarctic arine and fresh water sediments. J. Applied Bacteriol., 41: 75-80.
- 38. Fowler, V.J., N. Pfennig, W. Schubert and E. Stackbrandt, 1984. Towards a phylogeny of phototrophic purple sulfur bacteria-16S rRNA oligonucleotide cataloguing of 11 species of *Chromatiaceae*. Arch. Microbiol., 139: 382-387.
- Eichler, B. and N. Pfennig, 1988. A new purple sulfur bacterium from stratified freshwater lakes, *Amoebobacter purpureus* sp. nov. Arch. Microbiol., 149: 395-400.
- 40. Imhoff, J.F. 2001b. Transfer of *Pfennigia purpurea* Tindall 1999 (*Amoebobacter purpureus* Eichler and Pfennig 1988) to the genus *Lamprocystis* as *Lamprocystis purpurea* comb. nov. Int. J. Syst. Evol. Microbiol., 51: 1699-1701.
- 41. Tindall, B.J., 1999. Transfer of Amoebobacter purpureus to the genus Pfennigia gen. nov., as Pfennigia purpurea comb. nov., on the basis of the illegitimate proposal to make Amoebobacter purpureus the type species of the genus Amoebobacter. Int. J. Syst. Bacteriol., 49: 1307-1308.
- Winogradsky, S., 1888. Beitra ge zur Morphologie und Physiologie der Bakterien. Heft 1. Zur Morphologie und Physiologie der Schwefelbakterien, Arthur Felix, Leipzig, pp. 1-120.
- 43. Schmidt, K., N. Pfennigand S. Liaaen-Jansen, 1965. Cartenoids of *hiorhodaceae*. IV The carotenoids composition of 25 pure isolates. Arch Microbiol., 52: 132-146.